## Drug selection for transgenesis in Caenorhabditis species

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## Outline

- Why drug selection?
- Available drug selection systems
- Which drug is best?
- Dual drug selection for bombardment & its use in non-elegans species

## Why drug selection?





#### Rapid and easy (for bombardment)

- unc-119 worms tricky to grow in large numbers
- Selection by starvation (several weeks) vs 4 day drug selection

#### Universal (dominant marker)

- No requirement for specific genetic background
- Can be used for any species and any strain

## Why drug selection?

#### Other dominant markers (rol-6, fluorescent)

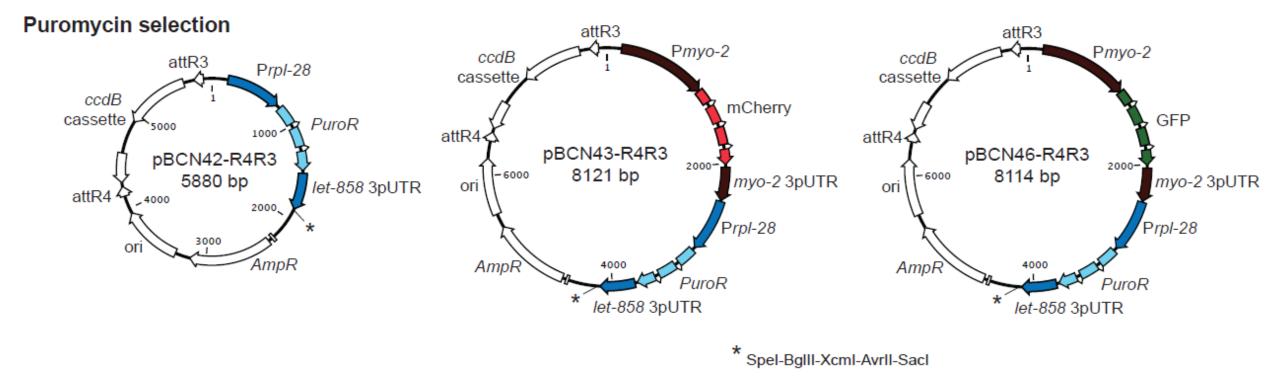
- $^\circ$  Easy visual tracking of transgene for crosses  $\checkmark$
- Difficult to select

#### **Drug** selection

• Can easily select for rare events without specialist equipment

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## Drug & fluorescence combo



PuroR = puromycin N-acetyl-transferase gene from Streptomyces bacteria + synthetic introns

### Drug selection systems available

#### Puromycin (Ben Lehner Lab)

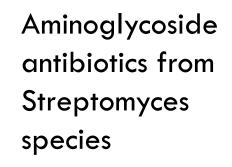
- Nat Methods, 2010. 7(9):725-7
- Nat Methods, 2012. 9(2):118-9

#### Neomycin/G418 (Denis Dupuy Lab)

• Nat Methods, 2010. 7(9):721-3

#### Hygromycin B (Jason Chin Lab)

• J Am Chem Soc, 2011. 133(36):14196-9 Expanding the genetic code of an animal.



Inhibit ribosome activity

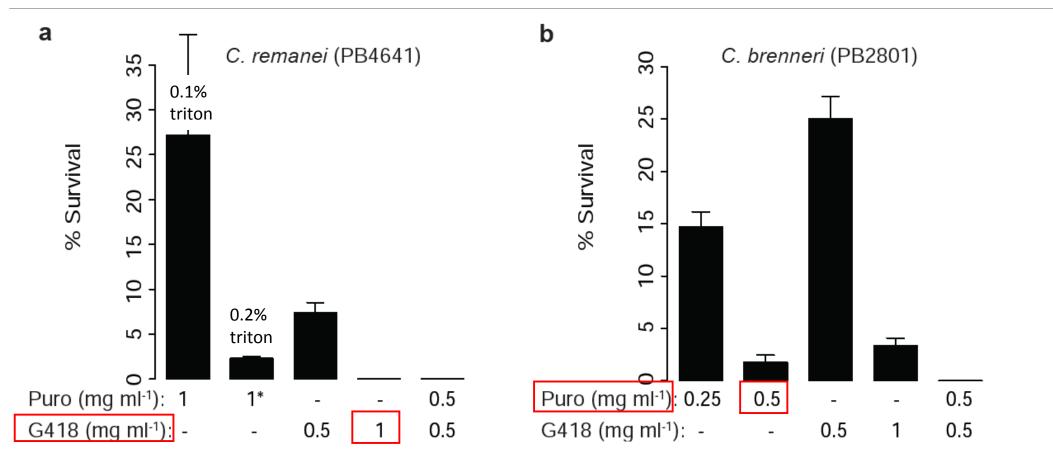
## Which drug is best?

Depends on the application:

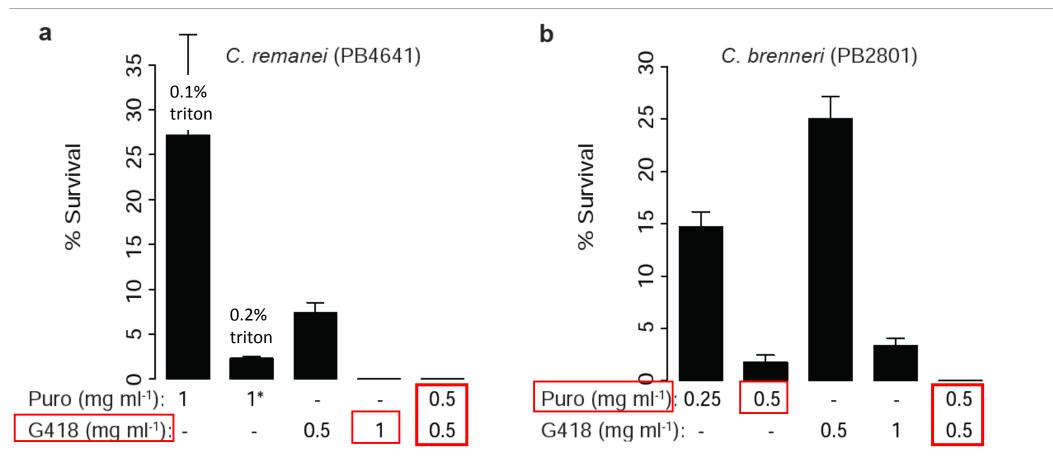
Drug	Price per Bombardment	Selection on plates	Liquid selection	Bombardment C. elegans	Bombardment other species
Puromycin (+triton)	7.6-26 USD	* (because of price)	***	***	Depends on species
G418	0.42-1 USD	***	**	***	Depends on species
Hygromycin B	0.68-3 USD	***	Ś	***	Ś
Puromycin + G418	4-13.5 USD	NA	***	***	***

Suppliers: Sigma, ForMedia, InvivoGen (?)

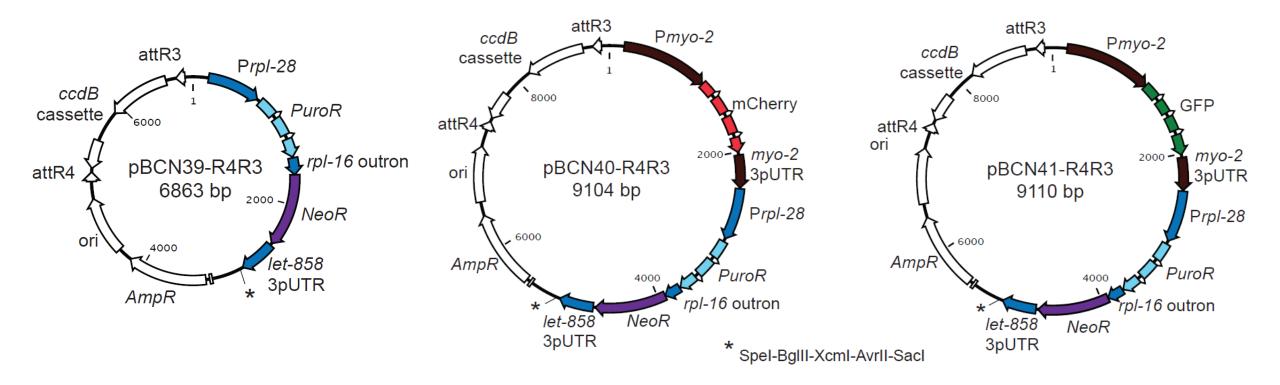
### Species variation in drug resistance



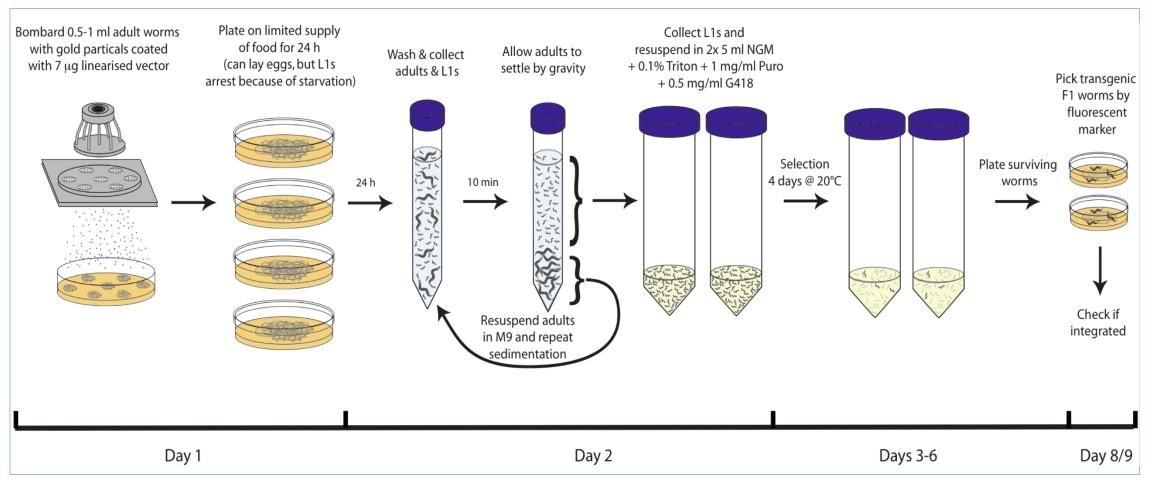
### Species variation in drug resistance

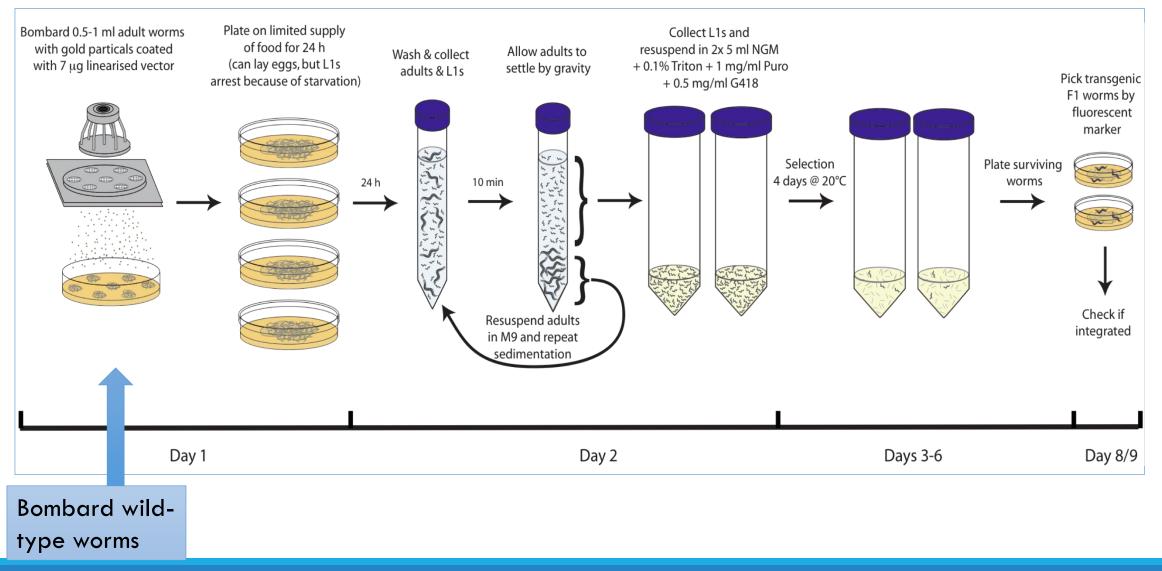


### Dual drug selection for bombardment



PuroR = puromycin N-acetyl-transferase gene from Streptomyces bacteria + synthetic introns



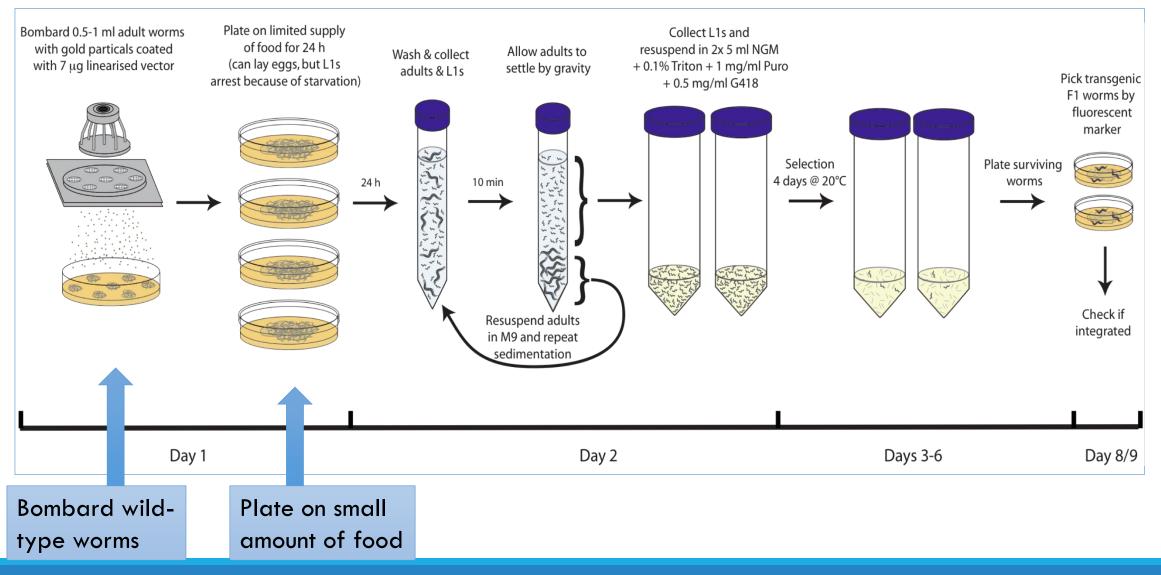


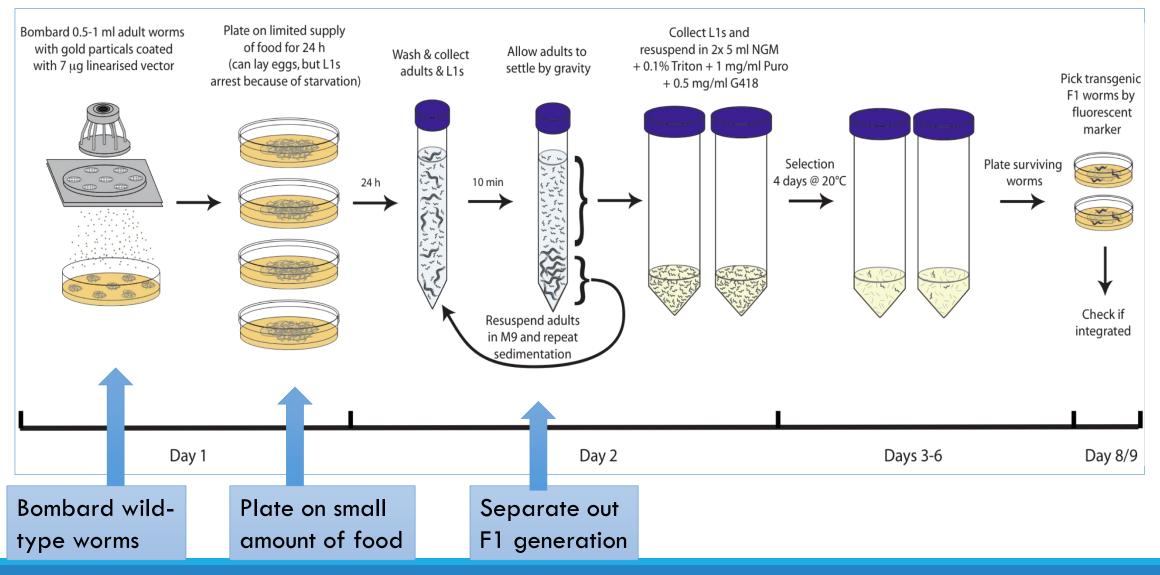
### Bombardment tips

- Use >1 ml of worms (~100,000 worms for N2):
- C. elegans:
  - 5 egg yolk plates with 20,000 worms/plate
- C. briggsae (smaller), C. remanei and C. brenneri (only 50% will contain eggs):

7-8 egg yolk plates with 20,000 worms/plate

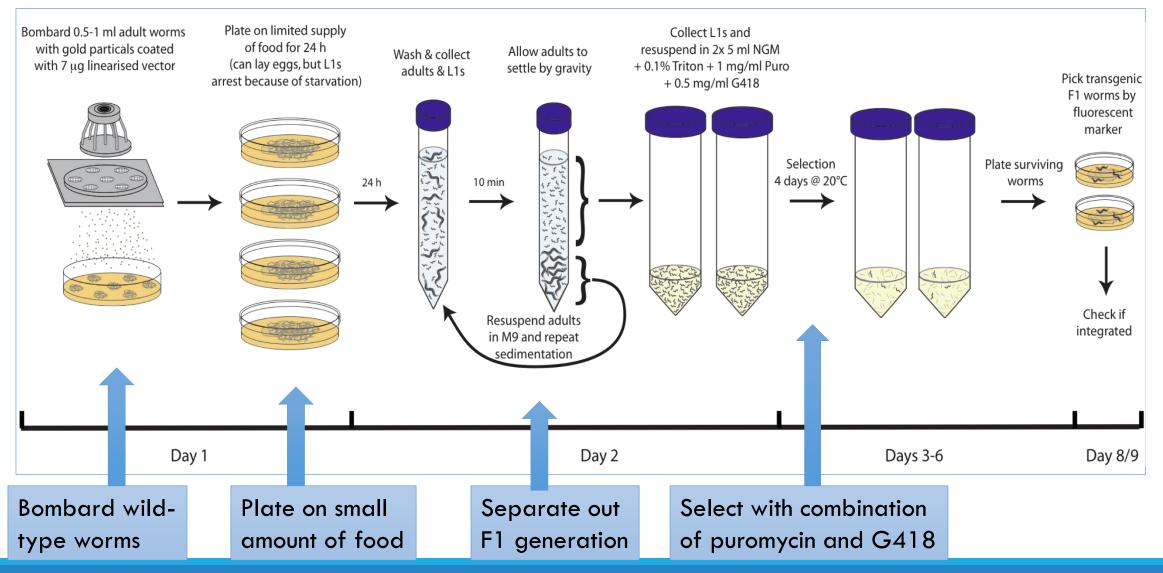
C. remanei and C. brenneri tend to burrow into plates : 2.5-3% NGM agar plates to make egg yolk plates





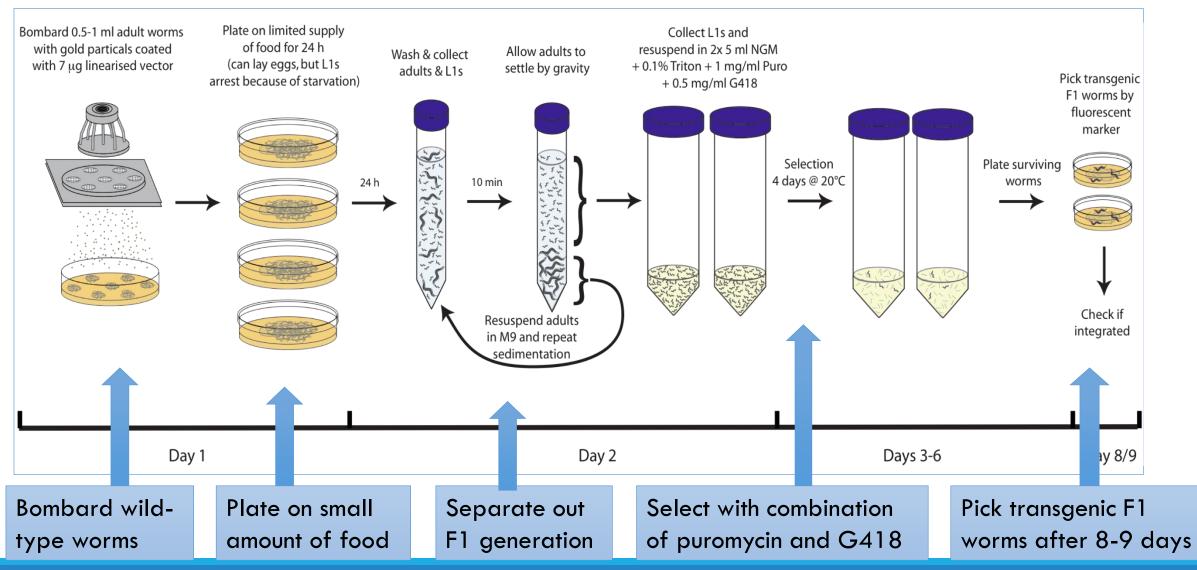
### Washing off worms and separating by gravity 2x

Very few L1s after 24h (slower development of some species): After separation of L1s, bleach adults and allow embryos to hatch overnight before selection.



### Selection with puromycin + G418

- Total volume 10-15 ml (~100 worms/µl)
- Good aeration: divide between 2x 50 ml conical tubes laid on their side (shaking incubator)



### Post bombardment

- Plate L1s on 2x 60mm NGM plates
- Integrated lines may have weak expression!! (low copy number)
- Usually get 10-1000 worms surviving, several extrachromosomal lines and 1-2 integrated
- Use fluorescent marker to pick transgenic worms uniform expression in whole pharynx

### Most common mistakes

- Not enough worms for bombardment
- Not enough perseverance scanning for weakly expressing transgenic worms

## Summary

- Drug selection is independent of genetic background
- Can be easily used with a variety of Caenorhabditis species and strains
- Single drug selection CAN be used for bombardment (1 mg/ml)
- Dual drug selection more robust to differences in resistance between species
- Vectors available from Addgene

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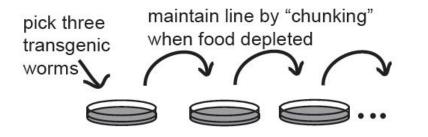
PEOPLE

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#### Maintaining non-integrated strains on plates



BCN6005 after 3 "generations" without selection



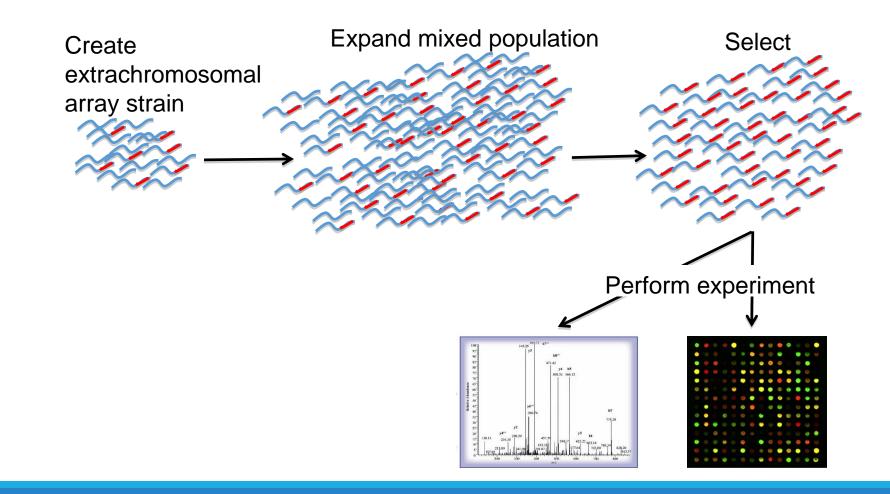


BCN6005 after 8 "generations" with selection



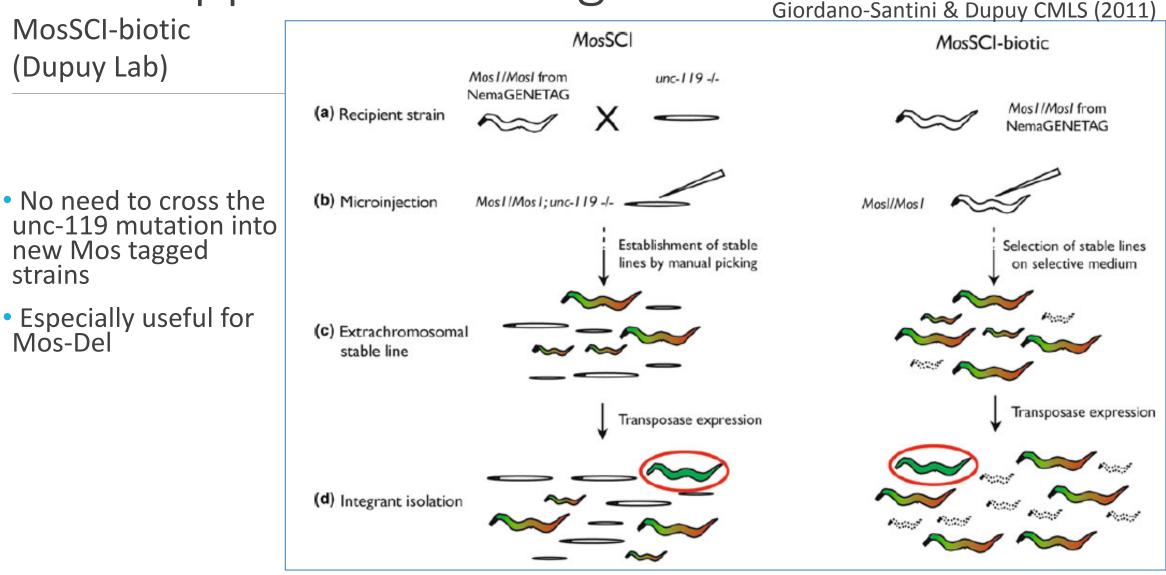
NGM agar + 0.5 mg ml<sup>-1</sup> puromycin

Obtaining large populations of transgenic worms from non-integrated strains



Generating integrated lines:

- MosSCI (Dupuy Lab)
- Biolistic Bombardment (Lehner Lab)



#### (Lehner Lab)

